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Synthesis Of Silicon Precursors Of Modified Oligonucleotides

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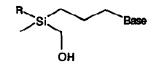
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Abstract: The synthesis of four silicon nucleoside analogues for use as modified antisense oligonucleotide precursors, is described.

In the field of antiviral or anticancer chemotherapy, the use of antisense oligonucleotides is increasingly promising 1, 2. Expression of viral proteins can be inhibited by the base-pairing of oligonucleotide sequences complementary to the coding sense strand of viral mRNA 3, 4. Such strands can be made of a small number of nucleotides (< 20) with a high specificity 5, and can be synthesized with automated DNA synthesizers. Nevertheless, there are limiting factors for the therapeutic use of natural antisense oligonucleotides such as degradation by nucleases, and low cellular uptake due to their strong anionic nature. To overcome these difficulties, the synthesis of oligonucleotides chemically modified in either the phosphodiester or ribofuranose components has been undertaken (see ref. 1 for a review). Recently, modified oligonucleotides have been synthesized in which the ribofuranose moiety has been replaced by 2-aminoethyl glycine units 6.

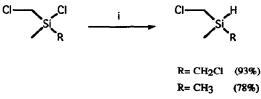
Our strategy, is to incorporate within an antisense oligonucleotide, nucleoside analogues consisting of organosilicon monomeric units. We expect from the presence of silicon, to increase the overall lipophily and to exploit modification of the fourth valency on the silicon atom for various purposes (improved solubility, cellular diffusion, ...). We report here the synthesis of four such organosilicon compounds (1a-d) bearing thymine and adenine nucleobases and one or two hydroxymethyl groups attached to the silicon atom via the phosphoramidite protocol for the solid phase polymer synthesis.



1 a :	R=CH2OH	Base= Thymine
1b:	R= Me	Base= Thymine
1c:	R=CH2OH	Base= Adenine
1d:	R= Me	Base= Adenine

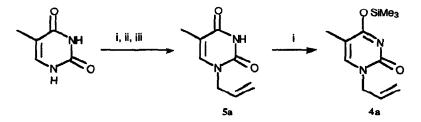
The key step of this synthesis lies in a hydrosilylation reaction between an N-allyl nucleobase and a

suitable silane prepared by reduction of the corresponding chlorosilane 7 (Scheme 1).



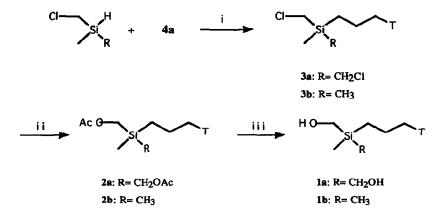
Scheme 1 - i: LiAlH4, Et2O, 0°C

For the preparation of the target molecules 1a and 1b, thymine was derivatized as its bis(trimethylsilyloxy) derivative, alkylated with allyl bromide to give a mixture of N-1-allylthymine 8 5a (65%) and N.N-1,3-bis(allyl)thymine (15%) as the only detected by-product. Finally, we found more convenient to protect 5a as its trimethylsilyl ether 4a since the subsequent hydrosilylation reaction was more efficient in the absence of the labile N-3 proton (Scheme 2).



Scheme 2 - i: HMDS, (NH4)2SO4; ii: allyl bromide/DMF, 80°C, 72h; iii: H2O, 0°C.

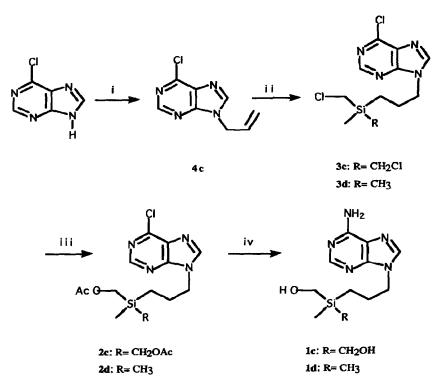
Thus, 4a was reacted with bis(chloromethyl)methylsilane or bis(methyl)chloromethylsilane with chloroplatinic acid as catalyst to yield 3a (35%) and 3b (20%) respectively (Scheme 3).



Scheme 3 - i: 1) H2PtCl6, THF 2) H2O; ii: AcONa, DMF, iii: 1) K2CO3, MeOH 2) HCl 1N.

The other steps of the synthesis involved the nucleophilic displacement of the chloromethyl compounds 3a and 3b with sodium acetate to give the ester precursors 2a (72%) and 2b (54%). Subsequent saponification (MeOH, K₂CO₃) afforded 1a (76%)⁹ and 1b (60%)¹⁰.

Compounds 1c and 1d (base= adenine) were prepared as outlined below (Scheme 4).



Scheme 4 - i: 1) NaH/DMF 2) allyl bromide, DMF; ii: 1) ClCH₂(Me)SiH or (ClCH₂)₂(Me)SiH, H₂PtCl₆, THF 2) H₂O; iii: AcONa/DMF; iv: NH₃/MeOH.

We started from the readily available 6-chloropurine ¹¹, as its allyl derivative 4c gave better yields of hydrosilylation products than allyladenine we used in our pilot studies (average yield 60% vs. 3%). The sodium salt of 6-chloropurine was alkylated with allyl bromide to afford 4c (65%) which, upon hydrosilylation, yielded 3c (57%) and 3d (17%). Compounds 3c and 3d were acetoxylated to give 2c (66%) and 2d (not isolated) which were further reacted with methanolic ammonia to yield the adenyl derivatives 1c (65%)¹² and 1d (50% from 3d)¹³.

The synthesis of poly-T and poly-A oligodeoxynucleotides including one or several **la-d** monomeric units is currently underway, and our results will be reported later.

Acknowledgments

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- Satisfactory spectroscopic data (FT-IR, 200 MHz ¹H and 50 MHz ¹³C NMR) were obtained for all compounds. 1a, mp= 116 °C, ¹H NMR (CD₃OD), δ (ppm) 0.07 (s, 3H, SiMe), 0.62-0.71 (m, 2H, SiCH₂), 1.66-1.82 (m, 2H, Si-C-CH₂), 1.85 (d, 3H, ⁴J_{Me,H-6}= 1 Hz, Me of thymine), 3.43 (s, 4H, 2 CH₂OH), 3.69 (t, 2H, J= 7.3 Hz, CH₂N), 7.42 (d, 1H, H-6); ¹³C NMR (CD₃OD), δ (ppm) -8.25 (SiMe), 8.34 (SiCH₂), 12.20 (Me of thymine), 24.17 (Si-C-C), 52.13 (CH₂N), 52.50 (CH₂OH), 110.86 (C-5), 143.22 (C-6), 152.82 (C-2), 166.76 (C-4).
- Compound 1b, mp= 138 °C, ¹H NMR (CDCl₃), δ (ppm) -0.01 (s, 6H, SiMe₂), 0.49-0.58 (m, 2H, SiCH₂), 1.58-1.74 (m, 2H, Si-C-CH₂), 1.85 (d, 3H, ⁴J_{Me,H-6}= 1 Hz, Me of thymine). 2.16 (large s, exchanged with D₂O, 1H, OH), 3.35 (s, 2H, CH₂OH), 3.62 (t, 2H, CH₂N), 6.99 (d, 1H, H-6), 9.94 (large s, 1H, NH); ¹³C NMR (CDCl₃), δ (ppm) -5.09 (SiMe₂), 10.42 (Si-C), 12.39 (Me of thymine), 23.61 (Si-C-C), 51.48 (CH₂N), 54.96 (CH₂OH), 110.57 (C-5), 140.62 (C-6), 150.96 (C-2), 164.26 (C-4).
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- Compound 1c, mp= 146 °C, ¹H NMR (DMSO-*d*₆), δ (ppm) -0.06 (s, 3H, SiMe), 0.46-0.54 (m, 2H, SiCH₂), 1.79-1.87 (m, 2H, Si-C-CH₂), 3.17 (s, 4H, 2 CH₂OH), 3.96 (large s, exchanged with D₂O, 2H, 2 OH), 4.08 (t, 2H, CH₂N), 7.29 (s, exchanged with D₂O, 2H, NH₂), 8.13 (2s, 2H, H-2 and H-8); ¹³C NMR (DMSO-*d*₆), δ (ppm) -7.93 (SiMe), 7.74 (Si-C), 24.11 (Si-C-C), 45.91 (CH₂N), 50.22 (CH₂OH), 118.74 (C-5), 141.11 (C-8), 149.54 (C-4), 151.84 (C-2), 155.51 (C-6).
- Compound 1d, mp= 166-170 °C, ¹H NMR (CDCl₃), δ (ppm) 0.04 (s, 6H, SiMe₂), 0.55-0.63 (m, 2H, SiCH₂), 1.86-2.01 (m, 3H, Si-C-CH₂ + OH), 3.40 (s, 2H, CH₂OH), 4.19 (t, 2H, J= 7 Hz, CH₂N), 5.76 (large s, exchanged with D₂O, 2H, NH₂), 7.79 (s, 1H, H-8), 8.34 (s, 1H, H-2); ¹³C NMR (CDCl₃), δ (ppm) -5.17 (SiMe), 10.56 (Si-C), 24.74 (Si-C-C), 46.69 (CH₂N), 54.40 (CH₂OH), 140.41 (C-8), 152.84 (C-2), 155.33 (C-6).

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